

Review article: the many potential roles of intestinal serotonin (5-hydroxytryptamine, 5-HT) signalling in inflammatory bowel disease

M. D. Coates¹  | I. Tekin² | K. E. Vrana³ | G. M. Mawe⁴

¹Department of Medicine, Division of Gastroenterology & Hepatology, Penn State Hershey Medical Center, Hershey, PA, USA

²Neuroscience Institute, University of California at Santa Barbara, Santa Barbara, CA, USA

³Department of Pharmacology, Penn State College of Medicine, Hershey, PA, USA

⁴Department of Neurological Sciences, University of Vermont College of Medicine, Burlington, VT, USA

Correspondence

Dr MD Coates, Division of Gastroenterology and Hepatology, Penn State University Hershey Medical Center, Hershey PA, USA. Email: mcoates@pennstatehealth.psu.edu

Funding information

The preparation of this work was supported in part by The Peter and Marsha Carlino Early Career Professorship (MDC), The Margo E. Walrath Career Professorship (MDC), Penn State Elliot S. Vesell Professorship endowment (KEV), and NIH grant DK 62267 (GMM).

Summary

Background: Serotonin (5-hydroxytryptamine, 5-HT) is an important mediator of every major gut-related function. Recent investigations also suggest that 5-HT can influence the development and severity of inflammation within the gut, particularly in the setting of inflammatory bowel disease (IBD).

Aim: To review the roles that the intestinal serotonin signalling system plays in gut function, with a specific focus on IBD.

Methods: We reviewed manuscripts from 1952 to 2017 that investigated and discussed roles for 5-HT signalling in gastrointestinal function and IBD, as well as the influence of inflammation on 5-HT signalling elements within the gut.

Results: Inflammation appears to affect every major element of intestinal 5-HT signalling, including 5-HT synthesis, release, receptor expression and reuptake capacity. Importantly, many studies (most utilising animal models) also demonstrate that modulation of selective serotonergic receptors (via agonism of 5-HT₄R and antagonism of 5-HT₃R) or 5-HT signal termination (via serotonin reuptake inhibitors) can alter the likelihood and severity of intestinal inflammation and/or its complicating symptoms. However, there are few human studies that have studied these relationships in a targeted manner.

Conclusions: Insights discussed in this review have strong potential to lead to new diagnostic and therapeutic tools to improve the management of IBD and other related disorders. Specifically, strategies that focus on modifying the activity of selective serotonin receptors and reuptake transporters in the gut could be effective for controlling disease activity and/or its associated symptoms. Further studies in humans are required, however, to more completely understand the pathophysiological mechanisms underlying the roles of 5-HT in this setting.

1 | INTRODUCTION

Serotonin (5-hydroxytryptamine, 5-HT) is a signalling molecule that plays critical roles in a large and varied number of human physiological functions, ranging from modifying mood to blood pressure control. Nowhere is the influence of 5-HT more ubiquitous, however, than in the gastrointestinal tract. The gastrointestinal mucosa harbours the largest store of 5-HT in the body.¹ Specifically, 5-HT is stored in granules and released from enterochromaffin (EC) cells residing in the lining of the intestinal crypts. Once released into the underlying lamina propria, 5-HT interacts with serotonergic receptors to affect a variety of functions in the gut before being removed by the serotonin selective reuptake transporter (SERT) (Figure 1). 5-HT influences every major function inherent to the gut, including motility, secretion, blood flow and sensation. Therefore, it is not surprising that when intestinal serotonergic signalling elements are disrupted (through disease processes, medications and/or inherent disorders of intestinal neuroendocrine function), these functions are impacted, resulting in myriad problematic symptoms that include abdominal pain, diarrhoea and/or constipation. It has been demonstrated, in both animal and human studies, that serotonergic signalling elements can have a major impact on inflammation development and severity within the gut, particularly in inflammatory bowel disease (IBD).^{2,3} There is strong evidence that modulation of a variety of 5-HT signalling components (including proteins related to 5-HT production and signal termination, as well as 5-HT-responsive receptors) can alter the risk of intestinal inflammation and its complications. In this article, we review the roles of 5-HT in normal intestinal functions, as well as more recent evidence demonstrating the impact that 5-HT signalling elements and inflammation within the gut have on each other. We propose the concepts that (1) the intestinal 5-HT signalling system exerts significant influence on the course of IBD and its symptomatic sequelae and (2) should be studied further as a target for disease modulation.

2 | SEROTONIN SIGNALLING AND FUNCTION WITHIN THE GUT

A common feature of visceral sensory, secretory, vasodilatory and motility pathways is that they can all be initiated by the action of 5-HT. In the gut, 5-HT can be found in specific enteric neurons, primarily descending interneurons in the myenteric plexus, but it is also produced, stored and secreted by EC cells in the mucosa. In fact, the vast majority of the body's 5-HT is located in EC cells.⁴ They represent a specialised subset of enteroendocrine cells, that are themselves one of the four main descendants of endodermally derived GI epithelial stem cells.⁵ The first described isoform of tryptophan hydroxylase (TPH-1) is the rate-limiting enzyme involved in the majority of 5-HT synthesis within the periphery, including the gastrointestinal tract, where it is responsible for 5-HT synthesis within the EC cells.⁶ Once synthesised, 5-HT is stored in granules within the EC cell until released by an appropriate

stimulus. The more recently discovered TPH-2 enzyme is responsible for 5-HT synthesis in enteric neurons (and neuronal 5-HT synthesis, in general).

Bulbring and colleagues revealed that intestinal mucosa releases 5-HT when mechanically stimulated and they provided evidence suggesting that EC cells are responsible for this release.⁷⁻⁹ Subsequent studies demonstrated that a variety of alterations in GI luminal conditions, including acidity, pressure and increased nutrient concentration, induce intestinal mucosa to release 5-HT.¹⁰⁻¹³ While other neuroactive compounds, such as cholecystekinin (CCK) and motilin, are released by luminal stimulation,^{14,15} it is 5-HT that seems to play a particularly critical role in sensory perception, secretion and peristalsis. The 5-HT released from EC cells is important because it represents the earliest intercellular event in the transduction of mucosal stimuli that initiates the neuronal reactions responsible for visceral sensation and peristalsis.¹⁶ It is worth noting, however, that Spencer et al. have demonstrated that peristalsis can be initiated and carried out in the absence of mucosal 5-HT.¹⁷ Therefore, it seems that 5-HT is not essential for the activation of peristalsis.^{18,19}

After release from EC cells into the lamina propria, 5-HT can interact with serotonergic receptors on the projections of neurons intrinsic or extrinsic to the gut, as well as receptors on nearby epithelial and immune cells. These interactions can stimulate secretion of a variety of signalling mediators and can potentiate neurons to make them more likely to respond to another stimulus or directly activate an action potential.²⁰ The influence that 5-HT has on a particular cell's activity varies depending upon the prevalent receptors. Seven different classes of 5-HT receptor (5-HT₁-5-HT₇) have been identified, using their structural and transductional characteristics, and there are currently at least 13 distinct human subtypes²¹ (Table 1). A variety of 5-HT-specific receptors have been identified in tissues of the gut, including on neurons as well as smooth muscle, epithelial and EC cells. For example, immunocytochemical studies have demonstrated that 5-HT₃ receptors are expressed on the subepithelial terminals of extrinsic sensory neurons.^{4,22,23} Electrophysiological studies show that when these neurons are exposed to a 5-HT₃-selective antagonist, such as alosetron, the normally robust response they have to 5-HT stimulation is attenuated.²⁴ In addition, 5-HT_{2A} receptors located on gut smooth muscle cells can stimulate muscle contraction when activated by 5-HT and thereby indirectly activate 5-HT-sensitive mechanosensitive nerve fibres in the gut wall.²⁵ Intrinsic neurons involved with GI motility utilise a different combination of serotonin receptors. In vitro studies, using isolated guinea pig intestine, have revealed that peristaltic activity is significantly diminished when an antagonist to the 5-HT₇ receptor (such as 5-HTP-DP) is applied alone or when antagonists or an antagonist to the 5-HT₃ and 5-HT₄ receptors together (eg, tropisetron) are applied to the tissue.^{4,16,26} As previously indicated, recent studies have also demonstrated that several cell types associated with immunological function, including antigen presenting cells, B cells, T cells, eosinophils, basophils and mast cells, express a variety of serotonergic receptors (such as 5-HT_{1A}, 5-HT_{1B}, 5-HT_{2A}, 5-HT_{2B}, 5-HT₄ and

FIGURE 1 Normal Intestinal Serotonin Signaling Cycle. (A) An appropriate stimulus (acidity, shear force) triggers 5-HT release from enterochromaffin (EC) cells into the lamina propria. (B) 5-HT interacts with serotonin-specific receptors on neurites and other mucosal cells expressing serotonergic receptors. This results in activation of sensory, motor and interneurons associated with the gut. (C) 5-HT is taken up by the selective serotonin reuptake transporter (SERT), found throughout the epithelium, thereby terminating its receptor stimulation

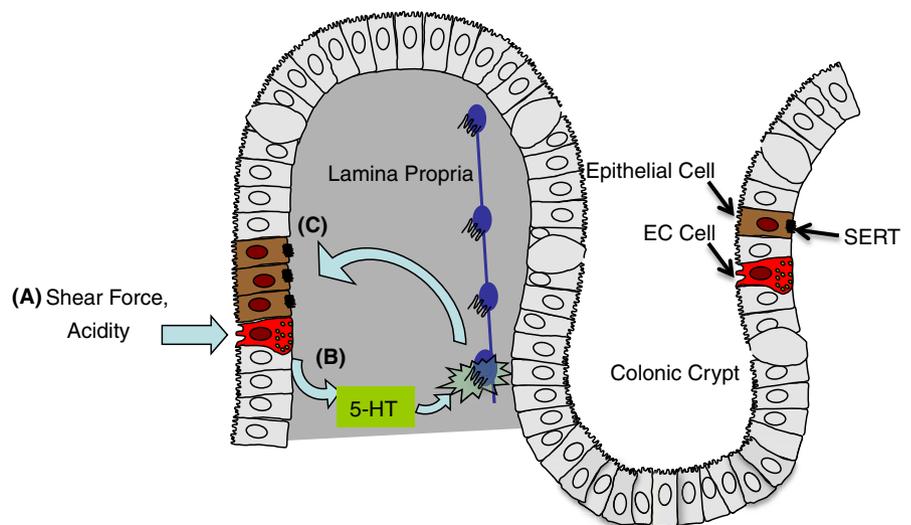


TABLE 1 5-HT receptors and their general function

Serotonin receptor type	Location	Function
5-HT _{1A}	CNS GI	Neuronal hyperpolarisation Mast cell degranulation; Mediator release
5-HT _{1B}	CNS	Inhibits neurotransmitter release, CV effects
5-HT _{1D}	CNS	Inhibits neurotransmitter release, CV effects
5-HT _{1E}	CNS	
5-HT _{1F}	CNS	Integrates sensorimotor info with limbic function
5-HT _{2A}	CNS GI	Influences effects of psychostimulants Contracts gut smooth muscle
5-HT _{2B}	GI	Enhances response of colonic smooth muscle
5-HT _{2c}	CNS	Influences emotional state
5-HT ₃	CNS GI	Modulates release of several neurotransmitters Modulates motility and visceral pain transmission
5-HT ₄	CNS GI	Impacts memory, cognitive function, affect Contracts gut smooth muscle
5-HT _{5A}	CNS	Regulates affect, learning and memory, perception, neuroendocrine function
5-HT _{5B}	CNS	
5-HT ₆	CNS	Regulates affect
5-HT ₇	CNS GI	Regulates affect Relaxes gut smooth muscle

CNS, central nervous system; GI, gastrointestinal; CV, cardiovascular.

5-HT₇) and that modulation of these receptors can induce significant changes in immune activity.^{27,28}

As with neurochemical transmission, to achieve precise reflex control with a paracrine transmitter, it is necessary to have a highly efficient mechanism to terminate the signal. This is primarily accomplished by the reuptake of 5-HT from the lamina propria by the SERT and its subsequent intracellular degradation by type A monoamine oxidase (MAO-A) or repackaging into vesicles.²⁹⁻³¹ SERT is a

transporter protein with 12 transmembrane-spanning domains whose activity is sodium-dependent.³² It is a highly selective 5-HT transporter that is expressed by serotonergic neurons in the ENS,⁴ but also by essentially all intestinal epithelial cells.^{24,29,31,33} Transgenic mice lacking SERT frequently exhibit diarrhoea associated with watery stools interspersed with periods of constipation.³⁰ We also know that SERT is involved in normal gut function in humans because alterations in bowel habits (eg, diarrhoea) are the most common side effect of serotonin selective reuptake inhibitors (SSRIs).³⁴⁻³⁶ Importantly, SERT synthesis in the gut involves a transcriptional start site that is downstream of that used by neurons,³⁷ and so appears to be regulated differently compared to what occurs in neurons. Finally, 5-HT that is not transported into epithelial cells enters the blood stream where it circulates in platelets that also express SERT.

All of the factors described above (5-HT synthesis and storage, release, receptor interaction and reuptake) are critical elements within the intestinal serotonin signalling system. Understanding how each of these factors work independently and in a coordinated fashion within the gut wall is important to properly understand not only intestinal serotonergic function, but also healthy and pathological gut physiology, including what occurs with inflammatory disorders of the gastrointestinal tract.

3 | IBD EPIDEMIOLOGY AND CHALLENGES

Inflammatory bowel disease, including ulcerative colitis (UC) and Crohn's disease (CD), affects an estimated 3 million people 18 years of age or older (1.3% of the adult population) in the USA alone.^{38,39} Despite recent advancements in therapy, many patients struggle finding adequate and/or consistent control of their disease and/or symptoms. Forty per cent or more of all IBD patients are eventually found to be refractory or intolerant to even the most efficacious therapies.^{40,41} Many find adequate control, but the path to success is frequently arduous and costly and, even when achieved, an

estimated 40% still report problematic symptoms (including abdominal pain and/or bowel habit changes) that significantly impact their quality of life.^{42,43} Patients with resistant disease and/or persistent symptoms incur higher healthcare expenditures and are more likely to miss work and report poor quality of life.⁴⁴

The symptoms described above, as well as the inflammatory state associated with IBD, may be affected by a variety of disparate factors, including genetic alterations relating to immune and epithelial barrier function, myriad environmental factors including diet and the gut microbiome and inherent host immune factors. Another underappreciated potential set of factors, however, are alterations in neuroendocrine signalling within the gut. In fact, there are many signalling molecules that can influence either the inflammation or symptoms associated with IBD. However, as indicated above, none has more wide-ranging effects and capability in this regard than 5-HT and perhaps no other neuroendocrine signalling system has demonstrated such profound changes as those seen with intestinal serotonergic signalling in IBD.

4 | RELATIONSHIPS AMONG INTESTINAL SEROTONIN SIGNALLING, INFLAMMATION AND ASSOCIATED SYMPTOMS

Several animal and human studies have demonstrated profound changes in one or more elements of 5-HT signalling in the setting of IBD (Tables 2 and 3), as well as significant effects of intestinal serotonergic signalling on the nature of inflammation in the gut (Tables 4 and 5). There is also evidence that 5-HT may be critical for development of a variety of symptoms in IBD, including abdominal pain and diarrhoea (Table 6). Here, we review the findings of those investigations, based upon the serotonergic signalling component(s) involved, the study models used and the inflammatory and/or functional end points evaluated.

4.1 | 5-HT synthetic capacity, content and EC cell density

4.1.1 | Animal studies

Most studies involving intestinal inflammation in mouse, rat and guinea pig (including those involving use of dextran sodium sulfate [DSS], trinitrobenzene sulfonic acid [TNBS], dinitrobenzene sulfonic acid [DNBS], acetic acid [AA] and *Trichuris suris* [TS] administration into the colon) have demonstrated significant increases in colonic mucosal 5-HT content and EC cell density.⁴⁵⁻⁵¹ A separate model using TNBS to stimulate ileitis in guinea pigs also demonstrated EC cell hyperplasia in the small intestine,⁵² while a canine enteritis model demonstrated elevated 5-HT content and EC cell density.⁵³ Two murine studies involving targeted cytokine knockouts (IL-2 and IL-13) did not exhibit these changes.^{54,55} Of note, the investigations utilising non-murine models are important in part because mice have mast cells within their intestinal tract that contain 5-HT (while the

other models do not) and the relative prevalence of these cells may contribute to changes in mucosal 5-HT levels during states of intestinal inflammation.⁵⁶

What is particularly interesting is the apparent impact that 5-HT has on the inflammatory process and certain symptoms associated with IBD. Two previous studies, using rodent models of colitis, demonstrated that application of the 5-HT precursor 5-hydroxytryptophan (5-HTP) worsened inflammation in each case.^{55,57} Concordantly, other studies that limited 5-HT production in murine models of colitis through either "genetic ablation" and/or pharmacological blockade of TPH-1 found that the inflammatory responses were significantly delayed or subdued as a result.⁵⁸⁻⁶⁰ Interestingly, this effect appeared to be specific to 5-HT localised in mouse intestinal mucosa.⁶⁰ Beyond this, another study, using croton oil to induce colitis in mice, found that application of the 5-HT precursor 5-HTP could induce more diarrhoea in these animals.⁶¹

4.1.2 | Human cell line and tissue studies

As early as the 1960s, physician-investigators started reporting that serotonergic elements are altered in the intestines of individuals with IBD. Verity⁶² and Capurso and Friedmann⁶³ were the first to demonstrate that 5-HT content was diminished in UC. A subsequent study including UC and CD patients also demonstrated a drop in colonic mucosal 5-HT content.⁵⁶ However, at least three studies demonstrated a rise in 5-HT content and/or EC cell density compared to healthy controls.⁶⁴⁻⁶⁶ These differences, however, may be explained by disease severity. For example, investigations that included specific assessment of severe UC demonstrated significant drops in EC cell density and 5-HT content in patients with the worst degree of inflammation.^{33,63}

Serotonin modifies gut inflammation in humans as well. In a study utilising three different human intestinal epithelial cell lines (CCD, HT-29 and CaCo-2), 5-HT application was found to significantly increase production of reactive oxygen species and monocyte epithelial adhesion.⁵⁷

Serotonin synthetic capacity and prevalence can also play an important role in symptoms frequently associated with IBD, regardless of disease activity state. Patients with colonic CD who demonstrate persistent irritable bowel syndrome (IBS)-like symptoms exhibit increased levels of TPH-1 RNA in colonic mucosal biopsies.⁶⁷ Individuals with an ileal pouch anal anastomosis (IPAA) who report persistent lower abdominal discomfort and alterations in bowel habits without overt inflammatory changes have also been found to have increased EC cell density in pouch biopsies.⁶⁸

4.2 | 5-HT release

4.2.1 | Animal studies

Every animal model of colitis (performed in mice and guinea pigs) evaluating 5-HT release has demonstrated an increase in epithelial 5-HT secretion compared to control conditions.^{46,52,69}

TABLE 2 Impact of inflammation on intestinal mucosal serotonin signaling in animal models of IBD

Citation	Study design	5-HT content	EC cell density	5-HT release	5-HT receptor Expression/function	SERT expression/function
Oshima et al. ⁴⁵	Rat DSS colitis	↑	↑			
Qjan et al. ⁵⁴	Mouse IL-2 knockout		NC			
Linden et al. ⁴⁶	Guinea Pig colitis	↑	↑	↑		↓ (mRNA, IR)
O'Hara et al. ⁵²	Guinea Pig TNBS ileitis		↑	↑		↓ (IR)
Linden et al. ⁸⁴	Mouse TNBS colitis					↓ (mRNA, IR, uptake)
Magro et al. ⁵¹	Mouse TNBS colitis	↑				
O'Hara & Sharkey ⁴⁹	Guinea Pig TNBS colitis		↑			
Bertrand et al. ⁶⁹	Mouse DSS colitis		↑	↑		↓ (mRNA)
Haub et al. ⁹⁰	IL-10 knockout		↓			
Matsumoto et al. ⁷²	Mouse DSS colitis	↑			↑ (5-HT ₃ R), ↓ (5-HT ₄ R)	
Shajib et al. ⁵⁵	Mouse IL13 Knockout/DSS colitis	↓	↓			
Guseva et al. ⁸²	Mouse IL10 Knockout/DSS colitis				↑ (5-HT ₇ R)	
El-Salhy et al. ⁵⁰	Rat TNBS colitis		↑			
Tada et al. ⁸⁵	Mouse DSS colitis					↓ (mRNA)
Bailey et al. ⁵³	Dog enteropathy	↑	↑		NC (5-HT _{2B} R)	NC

↑=increased, ↓=decreased, NC=no change, IR=immunoreactivity.

No animal studies have thus far investigated the impact of 5-HT release modulation on risk or severity of inflammation in this setting.

4.2.2 | Human cell line and tissue studies

An isolated EC cell line exposed to the inflammatory mediators IL-1-beta and LPS, exhibited increases in 5-HT release,⁷⁰ while the sole investigation looking at human IBD (UC) tissue found that 5-HT release was similar to that seen in tissue from healthy controls.³³ In addition, in a study of 85 patients with IBD who had undergone an IPAA, patients with the most severe endoscopy subscore on the Pouchitis Disease Activity Index demonstrated the highest levels of 5-HT in their blood.⁷¹

4.3 | 5-HT receptor expression and function

4.3.1 | Animal studies

There is surprisingly little published research that systematically describes 5-HT receptor distribution within the gut wall in the setting of intestinal inflammation. There is one animal study, which utilised a DSS colitis model in mice, that demonstrated an increase in 5-HT₃R and a decrease in 5-HT₄R within the colonic wall.⁷²

There is a growing body of literature, however, describing the influence that intestinal serotonergic receptors can exert on inflammation within the gut. Blocking 5-HT_{1A}R activity worsens TNBS-induced colitis in mice while stimulating the 5-HT_{1A}R or inhibiting the 5-HT_{2A}R reduces the severity of inflammation.⁷³ Conversely, 5-HT_{2A}R agonism created a "super-potent" reduction in pro-inflammatory markers in the small intestine of rats.⁷⁴ In the case of 5-HT₃R, two other studies incorporating mouse and rat AA colitis models

demonstrated that its antagonism (using granisetron and tropisetron, respectively) reduced inflammatory severity.^{75,76} Studies of 5-HT₄R agonism using animal colitis models suggested either no significant impact on inflammation^{73,77} or a protective effect of this receptor against inflammation.⁷⁸ Of note, Spohn et al. demonstrated that 5-HT₄R stimulation via enema administration had a protective effect against murine intestinal inflammation, but it was not protective when delivered by intraperitoneal injection.⁷⁸ In addition, normal animals treated by enema with a 5-HT₄R antagonist (GR113808) developed signs of early inflammation. Increased enteric neuronal proliferation has also been demonstrated in mouse and guinea pig models of DSS-induced colitis through a 5-HT₄R-dependant mechanism.^{79,80} Finally, research evaluating intestinal 5-HT₇R function initially suggested that inflammation in DSS and DNBS colitis in mice could be reduced through pharmacological antagonism of this receptor or via genetic knock-out.⁸¹ However, a more recent study (utilising a similar murine colitis model) suggests that 5-HT₇R may play more of a protective role, reducing intestinal inflammatory activity when activated.⁸² In summary, it is clear that 5-HT receptors play important, though potentially differing, roles in the modulation of intestinal inflammation. Available data suggest that 5-HT_{1A}R, 5-HT_{2A}R, and 5-HT₄R activation is likely protective against inflammation in the gut while 5-HT₃R is more likely to worsen the inflammatory process. The evidence for the influence of 5-HT₇R is less clear.

5-HT receptor modulation could also have a significant impact on symptoms associated with gut inflammation. In the murine croton oil colitis study described above, it was found that application of a 5-HT₃R antagonist (ondansetron) could resolve the 5-HTP-induced diarrhoea exhibited in these mice.⁶¹ Another study, utilising DSS-induced colitis in mice, found that these animals not only

TABLE 3 Impact of inflammation on intestinal mucosal serotonin signaling in human studies of IBD

Citation	Disease type	5-HT content	EC cell density	5-HT release	5-HT receptor Expression/function	SERT expression/function
Verity ⁶¹	UC	↓				
Capurso & Friedmann ⁶³	UC and healthy controls	↓				
El-Salhy et al. ⁶⁴	UC and CD	↑	↑			
Stoyanova et al. ⁶⁵	UC, surgical specimens		↑			
Magro et al. ⁵⁶	UC and CD, surgical specimens	↓				
Coates et al. ³³	UC, mucosal biopsies	↓	↓ (severe)	NC		↓ (mRNA, IR)
Foley et al. ⁹³	CaCo-2 cells; IFN γ /TNF α Exp					↓ (mRNA, uptake)
Minderhoud et al. ⁶⁷	CD, ileal and colonic biopsies	↑ (TpH-1)	NC			
Kidd et al. ⁷⁰	EC Cell Isolates; IL-1b, LPS Exp			↑		
Latorre et al. ⁹⁴	CaCo-2 Cells	↑				↓ (low IL-10) ↑ (high IL-10)
Wang et al. ⁷¹	IPAA	↑ (severe)				
Guseva et al. ⁸²	CD				↑ (5-HT ₇ R)	
Sikander et al. ⁶⁶	Microscopic colitis, UC, Control	↑				
Tada et al. ⁸⁵	UC					↓
Yu et al. ⁸³	UC			↑ (Plasma 5-HT and 5-HIAA)	↑ (5-HT ₃ R, 5-HT ₅ R)	

↑=increased, ↓=decreased, NC=no change, IR=immunoreactivity, IPAA=ileal pouch anal anastomosis.

demonstrated visceral hypersensitivity, but that use of a 5-HT₃R antagonist (tropisetron) could attenuate this effect.⁷² More recently, Spohn et al.⁷⁸ demonstrated in a murine DSS colitis model that tegaserod (a 5-HT₄R agonist) improved intestinal motility while use of a 5-HT₄R antagonist (GR113808) in otherwise untreated mice resulted in the development of inflammation, dysmotility and obstruction in non-inflamed animals.

4.3.2 | Human cell line and tissue studies

In a study evaluating UC patients in remission, 5-HT₃R and 5-HT₅R transcript levels were both increased in colonic mucosal biopsies, while 5-HT₇R was unchanged.⁸³ 5-HT₇R density was also found to be increased in the setting of CD.⁸² It should be noted that these changes in receptor expression likely reflect receptors expressed by local cells in the mucosal layer, as opposed to neurons, because neuronal RNA is probably not collected in a routine colonic biopsy, as neuronal cell bodies are found either in deeper layers within the intestinal wall or outside of the gut all together.

4.4 | SERT expression and function

4.4.1 | Animal studies

Linden et al. were the first to demonstrate significant decreases in SERT expression in an animal (guinea pig) model of colitis.⁴⁶ Every subsequent study of SERT expression and/or function in animal models of colitis or enteritis has demonstrated this same finding.^{48,52,84-86}

Interestingly, use of SSRIs has been shown to be protective against inflammation in several animal models. Administration of fluoxetine in mice with DSS colitis resulted in reduction in inflammatory markers (eg, NF κ B) and histological inflammatory scores.⁸⁷ Intra-colonic fluvoxamine therapy reduced intestinal inflammatory activity and assessment scores in mice with AA-induced colitis.⁸⁸ Fluoxetine treatment of IL-10 deficient mice reduced intestinal inflammatory cell infiltration and reduced colitis scores.⁸⁹ However, other studies utilising SERT knockout (KO) mice demonstrated significantly worsened intestinal inflammation in the absence of the transporter.^{90,91}

4.4.2 | Human cell line and tissue studies

UC patients display markedly reduced SERT immunoreactivity and mRNA expression³³ and the latter has been observed in another cohort of UC patients with mild inflammation.⁸⁵ Similar findings were demonstrated in a cohort of diverticulitis patients.⁹² It is likely that interferon-gamma and TNF-alpha contribute to the down-regulation of SERT in colitis because SERT expression and function are decreased in CaCo-2 cells (which natively express SERT) when they are exposed to these pro-inflammatory cytokines.⁹³ In another study, interleukin-10 demonstrated the ability to up- and down-regulate SERT concentration and activity in CaCo-2 cells depending on the concentration of cytokine applied.⁹⁴

Several studies have also evaluated the impact of antidepressant medications on IBD symptom severity. Most of these investigations have found that the use of these antidepressants is associated with improved IBD symptom severity and/or disease scores.^{95,96} Although

TABLE 4 Effects of alterations in intestinal mucosal serotonin signaling on inflammation in animal models of IBD

Citation	Study design	Impact on inflammation
Shajib et al. ⁵⁵	Increasing 5-HT Stores (5-HTP) in mouse IL-13 knockout or DSS colitis	↑
Regmi et al. ⁵⁷	5-HT applied in rat TNBS colitis	↑
Ghia et al. ⁵⁸	TPH-1 knockout or inhibitor (PCPA) in mouse DSS, DNBS colitis	↓
	Increasing 5-HT stores (with 5-HTP)	↑
Margolis et al. ⁶⁰	Peripheral inhibition of TPH-1 in mouse TNBS colitis	↓
Kim et al. ⁵⁹	Peripheral TPH-1 inhibitor (Telotristat) in Mouse DSS or <i>Trichuris suris</i> colitis	↓
Rapalli et al. ⁷³	Selective 5-HT receptor agonism/antagonism in mouse TNBS colitis	
	5-HT _{1A} R agonist	↑
	5-HT _{1A} R antagonist	↓
	5-HT _{2A} R agonist	↓
	5-HT ₃ R, 5-HT ₄ R agonist, 5-HT ₇ R antagonist	NC
Nau et al. ⁷⁴	5-HT _{2A} R agonist in rat enteritis	↓
Mousavizadeh et al. ⁷⁶	5HT ₃ R antagonist (Tropisetron) in Rat AA Colitis	↓
Fakhfouri et al. ⁷⁵	5HT ₃ R antagonist (Granisetron) in mouse AA colitis	↓
Spohn et al. ⁷⁸	5-HT ₄ R agonist (Tegaserod) in rectal mouse DSS or TNBS colitis	↓
	5-HT ₄ R antagonist (GR113808)	↑
Koh et al. ⁸⁷	5-HT ₇ R knockout or inhibitor in mouse DSS colitis	↓
Kim et al. ⁸¹	5-HT ₇ R knockout or antagonist in mouse DSS or DNBS colitis	↓
Guseva et al. ⁸²	5-HT ₇ R knockout or antagonist in mouse IL-10 knockout or DSS colitis	↑
	5-HT ₇ R agonist	↓
Bischoff et al. ⁹¹	SERT knockout in mouse TNBS colitis	↑
Haub et al. ⁹⁰	SERT knockout in mouse IL-10 knockout	↑
Koh et al. ⁸⁷	Fluoxetine in mouse DSS colitis	↓
Minaiyan et al. ⁸⁸	Fluvoxamine in normal and reserpenised rats acetic acid colitis	↓
Koh et al. ⁸⁹	Fluoxetine in mouse IL-10 colitis	↓

↑=increased, ↓=decreased, NC=no change; 5-HTP=5-HT precursor.

TABLE 5 Effects of alterations in intestinal mucosal serotonin signaling on inflammation in human studies of IBD

Citation	Study design	Impact on inflammation
Regmi et al. ⁵⁷	Application of 5-HT to three human cell lines; measured ROS and MEA	↑
Fernandez-Banares et al. ⁹⁸	Retrospective assessment of risk for microscopic colitis in setting of SSRI use	↑
Bonderup et al. ⁹⁷	Case-control study assessing risk for microscopic colitis in setting of SSRI use	↑
Sikander et al. ⁶⁶	Presence of 5-HTTLPR s/s in microscopic colitis or UC (vs Control)	↓

↑=increased, ↓=decreased, ROS=reactive oxygen species, MEA=monocyte epithelial adhesion, s/s=homozygous for "short allele".

most of the agents in question could modulate 5-HT activity, none of these studies specifically evaluated selective serotonin reuptake inhibitor use alone in comparison to IBD disease activity. Although some of these assessments utilised questionnaires that targeted somatic symptoms (including abdominal pain and changes in bowel habits), it should be noted that psychiatric symptoms were a component for many of the disease activity indices utilised for these investigations. In addition, no human studies have utilised rigorous measures to objectively assess intestinal inflammatory status and/or other alimentary complications of IBD in relation to use of these agents. Of note, SSRI use has previously been linked to increased incidence of microscopic colitis, though the exact mechanism underlying this relationship is unclear.^{97,98}

5 | CONCLUSIONS

The intestinal serotonergic signalling system appears to play a significant and multi-faceted role in IBD. The studies described above demonstrate a profound effect of intestinal inflammation on mucosal serotonergic signalling. A variety of effects have been observed in different elements of intestinal 5-HT signalling in the setting of experimental colitis and IBD in humans, depending on the disease type or model, severity of disease and/or experimental design. Despite these variations, a few consistent themes have emerged from studies conducted to date. First, intestinal and serum 5-HT levels and synthetic capacity are altered in the setting of IBD. Second, mucosal 5-HT release is either unchanged or increased in these

TABLE 6 Impact of alterations in intestinal mucosal serotonin signaling on symptoms in animal and human models of IBD

Citation	Study design	Impact on symptoms
Pascual et al. ⁶¹	5-HTP application mouse croton oil colitis 5HT ₃ R antagonist (Ondansetron)	↑ (diarrhoea) ↓ (diarrhoea)
Matsumoto et al. ⁷²	5HT ₃ R antagonist (Tropisetron) in mouse DSS colitis	↓ (visceral hypersensitivity)
Spohn et al. ⁷⁸	5HT ₄ R agonist (Tegaserod) in mouse DSS colitis 5HT ₄ R antagonist (Tegaserod)	↓ (obstruction) ↑ (obstruction)
Minderhoud et al. ⁶⁷	Increased TpH-1 levels in colonic biopsies from CD colitis patients	↑ (IBS symptoms)
Shen et al. ⁶⁸	Increased EC cells in pouch biopsies from irritable pouch syndrome patients	↑ (IBS symptoms)

↑=increased, ↓=decreased; 5-HTP=5-HT precursor.

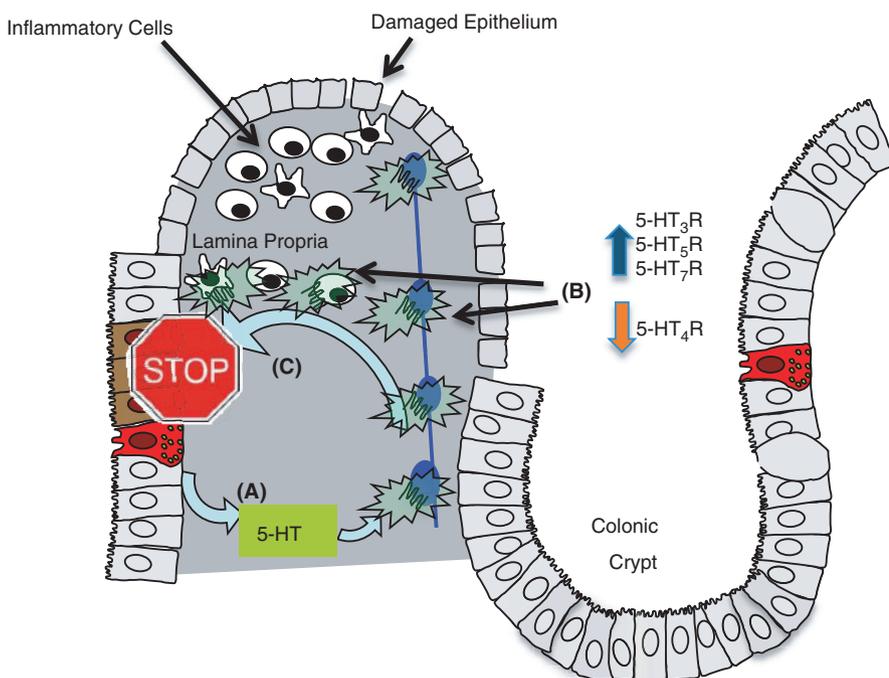


FIGURE 2 IBD impacts the intestinal serotonin signaling cycle. (A) 5-HT release appears to be the same or increased in the setting of inflammation. (B) Inflammation induces changes in serotonin-specific receptors (eg, increased 5-HT₃R, 5-HT₅R, 5-HT₇R and decreased 5-HT₄R) on neurites and other mucosal cells. (C) SERT prevalence is reduced in IBD, leading to reduced uptake (symbolised by the “stop sign”), increased 5-HT availability in the mucosa and variably modified activation of neurons and immune cells expressing serotonergic receptors

disorders. Third, SERT expression (and function based upon a few studies) is consistently and significantly diminished in IBD. Finally, 5-HT receptor expression (for a variety of subtypes) is altered in colitis. These findings suggest that intestinal 5-HT availability is increased overall in the setting of IBD and that many serotonergic signalling capabilities can be affected by inflammation (Figure 2).

Conversely, intestinal 5-HT and its receptors also appear to play a major role in modulating the development and intensity of inflammation. Empiric observations of populations associated with increased exposure to 5-HT and its metabolites (eg, carcinoid syndrome) suggest that there is an increased risk for IBD.^{99,100} It has also been demonstrated that several serotonergic components within the gut can enhance or diminish the inflammatory process in IBD and that these elements have the potential to be manipulated to significant effect. Increased 5-HT production and release along with the agonism of certain serotonergic receptors (eg, 5-HT₃R) appear to worsen the inflammatory process in IBD, while reduction in 5-HT availability and the activation of different receptors (eg, 5-HT_{1A}R, 5-HT₄R) appear to reduce or delay the same inflammatory process

(Figure 3). There are contradictory findings in animal models of colitis regarding the influence of SERT on disease activity and limited human data. SERT knockout models in rodents with colitis demonstrate worsening of the disease while SSRI administration in non-modified animals appeared to be protective against it. Of note, there is evidence that SSRIs may increase the risk for microscopic colitis and some preliminary data has suggested that SSRIs may actually put IBD patients at greater risk for disease flares⁹⁵ but there are no completed studies that have specifically evaluated this latter relationship. As previously stated, there are also no prior studies evaluating any anti-depressant in the setting of IBD that have utilised objective measures of inflammation (eg, endoscopy and/or histology scores).

Manipulations of 5-HT availability (including via use of agents that modulate SERT function) and certain serotonergic receptors (including 5-HT₃R, 5-HT₄R) also have the potential to impact risk for development of common problematic symptoms in IBD, including visceral hypersensitivity and alterations in bowel habits. Those symptoms directly account for a great deal of the morbidity and reduced quality of life associated with CD and UC, as well as the costs

(A) Pro-Inflammatory Effects

- 5-HT precursors (e.g., 5-HTP)
- 5-HT_{1A}R, 5-HT₃R agonists
- 5-HT₄R antagonists
- SERT Knockout

(B) Anti-Inflammatory Effects

- TPH Inhibitors (e.g., telotristat)
- 5-HT_{2A}R, 5-HT₄R agonist
- 5-HT_{1A}R, 5-HT₃R antagonist

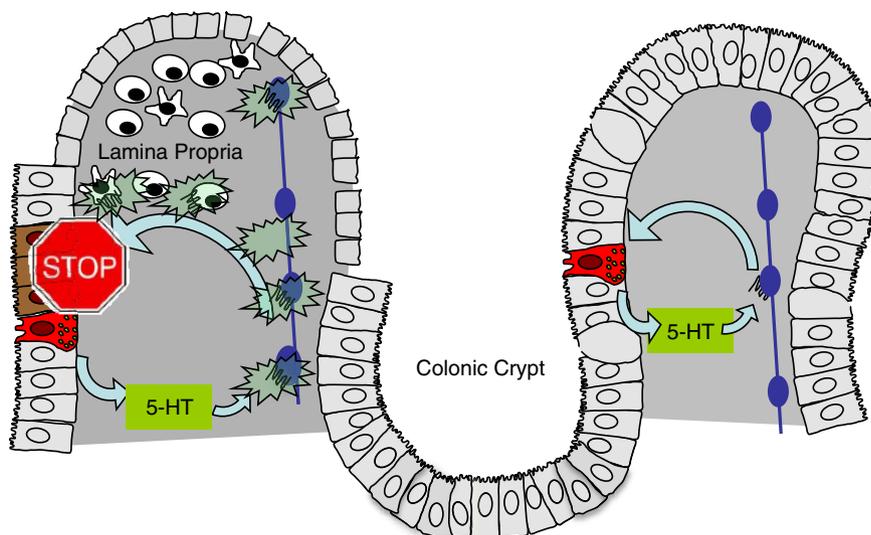


FIGURE 3 Serotonin signaling components impact inflammatory bowel disease activity and symptoms. (A) 5-HT (and its' precursors), 5-HT₃R agonists, 5-HT₄R antagonists and serotonin reuptake inhibitors promote inflammation and problematic symptoms in IBD. (B) Agents that decrease 5-HT production and release, 5-HT₄R agonists, 5-HT₃R antagonists reduce the likelihood of inflammation and symptoms

associated with increased healthcare resource utilisation and lost work hours. These findings closely mirror those found in investigations of IBS, another set of conditions associated with visceral hypersensitivity and altered bowel habits. Studies in both humans and animal models attempting to mimic IBS have demonstrated that increasing 5-HT availability and/or manipulating serotonergic receptor function can either induce or prevent development of visceral hypersensitivity and/or diarrhoea depending on the factors involved.^{33,61,101-105} Selective 5-HT receptor manipulation can also have a dramatic effect on visceral sensation and motility in IBS and animal models of IBS, as has been demonstrated with agonists and antagonists of the 5-HT₄ receptor.^{106,107} Although seemingly quite divergent from many phenotypic perspectives, IBD and IBS share many symptomatic similarities that may well share certain pathophysiological parallels. In fact, this has been hinted at by prior studies looking at 5-HT signalling factors in IBD and IBS populations simultaneously.³³

There remains a gap in our knowledge regarding the impact of alterations in intestinal serotonergic signalling on symptoms in IBD, particularly in humans (Table 6). There is also a dearth of information regarding the variability in every major element of intestinal 5-HT signalling with regard to intestinal region and disease severity. This is particularly true for CD. In addition, the expression and/or function of most 5-HT receptors have not been systematically evaluated in the context of either UC or CD. More comprehensive, longitudinal investigations utilising tissue samples throughout the intestinal tract during both quiescent and active phases of UC and CD would clarify the role that various serotonergic enzymes, transporters and receptors play in gastrointestinal signalling and in IBD.

In conclusion, the findings outlined in this review demonstrate the powerful influence that the neuroendocrine signalling mediator 5-HT can have on the development and perpetuation of IBD and associated symptoms and suggest new diagnostic and therapeutic opportunities to consider in the management of these disorders. In particular, more selective 5-HT receptor activation (eg, utilising 5-HT_{1A}R or 5-HT₄R agonists) and/or inhibition (eg, with 5-HT₃R antagonists), using targeted delivery strategies (eg, enemas or other topically applied approaches) to help enhance efficacy and reduce the potential for adverse side effects, may prove very effective as adjunctive therapy in the control of inflammation and symptoms of IBD. In addition, while judiciously selected and monitored serotonergically active antidepressant agents in these patients can help to address the mood disorders frequently associated with IBD and have the potential to mitigate problematic symptoms, these medications need to be evaluated more carefully as there is at least some evidence that they may increase risk for intestinal inflammation. Given the number of patients with IBD who continue to struggle achieving control of their disease and its associated symptoms, and the apparent influence of serotonergic signalling factors in this setting, further studies incorporating objective assessments of intestinal inflammation and 5-HT signalling in these disorders are certainly warranted. In time, it is likely that more serious consideration will need to be made to evaluate and intervene on serotonergic factors within the gut in the setting of IBD.

AUTHORSHIP

Guarantor of the article: Dr. Coates.

Author contributions: Dr Coates developed the idea for this article, performed literature searches, was the primary author and editor. Dr Tekin performed an independent literature review for this article and she helped to write and edit the manuscripts and tables. Dr. Vrana helped to write and edit every component of this submission. Dr. Mawe helped to write and edit each component of this submission. All authors have approved the final version of this submission.

ACKNOWLEDGEMENTS

Declaration of personal interests: None.

REFERENCES

- Erspamer V, Asero B. Identification of enteramine, the specific hormone of the enterochromaffin cell system, as 5-hydroxytryptamine. *Nature*. 1952;169:800-801.
- Goldner D, Margolis KG. Association of serotonin transporter promoter polymorphism (5HTTLPR) with microscopic colitis and ulcerative colitis: time to be AsERTive? *Dig Dis Sci*. 2015;60:819-821.
- Margolis KG, Gershon MD. Enteric neuronal regulation of intestinal inflammation. *Trends Neurosci*. 2016;39:614-624.
- Gershon MD. Review article: roles played by 5-hydroxytryptamine in the physiology of the bowel. *Aliment Pharmacol Ther* 1999;13 (Suppl. 2):15-30.
- Hocker M, Wiedenmann B. Molecular mechanisms of enteroendocrine differentiation. *Ann N Y Acad Sci*. 1998;859:160-174.
- McKinney J, Teigen K, Froystein NA, et al. Conformation of the substrate and pterin cofactor bound to human tryptophan hydroxylase. Important role of Phe313 in substrate specificity. *Biochemistry*. 2001;40:15591-15601.
- Bulbring E, Lin RC. The effect of intraluminal application of 5-hydroxytryptamine and 5-hydroxytryptophan on peristalsis; the local production of 5-HT and its release in relation to intraluminal pressure and propulsive activity. *J Physiol*. 1958;140:381-407.
- Bulbring E, Crema A. Observations concerning the action of 5-hydroxytryptamine on the peristaltic reflex. *Br J Pharmacol*. 1958;13:444-457.
- Bulbring E, Crema A. The release of 5-hydroxytryptamine in relation to pressure exerted on the intestinal mucosa. *J Physiol*. 1959;146:18-28.
- Kim M, Cooke HJ, Javed NH, Carey HV, Christofi F, Raybould HE. D-glucose releases 5-hydroxytryptamine from human BON cells as a model of enterochromaffin cells. *Gastroenterology*. 2001;121:1400-1406.
- Fukumoto S, Tatewaki M, Yamada T, et al. Short-chain fatty acids stimulate colonic transit via intraluminal 5-HT release in rats. *Am J Physiol Regul Integr Comp Physiol*. 2003;284:R1269-R1276.
- Yu PL, Fujimura M, Hayashi N, Nakamura T, Fujimiya M. Mechanisms in regulating the release of serotonin from the perfused rat stomach. *Am J Physiol Gastrointest Liver Physiol*. 2001;280:G1099-G1105.
- Kim M, Javed NH, Yu JG, Christofi F, Cooke HJ. Mechanical stimulation activates Galphaq signaling pathways and 5-hydroxytryptamine release from human carcinoid BON cells. *J Clin Invest*. 2001;108:1051-1059.
- Wang Y, Prpic V, Green GM, Reeve JR Jr, Liddle RA. Luminal CCK-releasing factor stimulates CCK release from human intestinal endocrine and STC-1 cells. *Am J Physiol Gastrointest Liver Physiol*. 2002;282:G16-G22.
- Goll R, Nielsen SH, Holst JJ. Regulation of motilin release from isolated perfused pig duodenum. *Digestion*. 1996;57:341-348.
- Jin JG, Foxx-Orenstein AE, Grider JR. Propulsion in guinea pig colon induced by 5-hydroxytryptamine (HT) via 5-HT4 and 5-HT3 receptors. *J Pharmacol Exp Ther*. 1999;288:93-97.
- Spencer NJ. Constitutively active 5-HT receptors: an explanation of how 5-HT antagonists inhibit gut motility in species where 5-HT is not an enteric neurotransmitter? *Front Cell Neurosci*. 2015;9:487.
- Spencer NJ, Sia TC, Brookes SJ, Costa M, Keating DJ. CrossTalk opposing view: 5-HT is not necessary for peristalsis. *J Physiol*. 2015;593:3229-3231.
- Smith TK, Gershon MD. CrossTalk proposal: 5-HT is necessary for peristalsis. *J Physiol*. 2015;593:3225-3227.
- Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. *Curr Opin Endocrinol Diabetes Obes*. 2013;20:14-21.
- Gershon MD. Review article: serotonin receptors and transporters – roles in normal and abnormal gastrointestinal motility. *Aliment Pharmacol Ther*. 2004;20(Suppl. 7):3-14.
- Read NW, Gwee KA. The importance of 5-hydroxytryptamine receptors in the gut. *Pharmacol Ther*. 1994;62:159-173.
- Raybould HE, Glatzle J, Robin C, et al. Expression of 5-HT3 receptors by extrinsic duodenal afferents contribute to intestinal inhibition of gastric emptying. *Am J Physiol Gastrointest Liver Physiol*. 2003;284:G367-G372.
- Bertrand PP, Kunze WA, Furness JB, Bornstein JC. The terminals of myenteric intrinsic primary afferent neurons of the guinea-pig ileum are excited by 5-hydroxytryptamine acting at 5-hydroxytryptamine-3 receptors. *Neuroscience*. 2000;101:459-469.
- Hillsley K, Grundy D. Sensitivity to 5-hydroxytryptamine in different afferent subpopulations within mesenteric nerves supplying the rat jejunum. *J Physiol*. 1998;509(Pt 3):717-727.
- Nagakura Y, Ito H, Kiso T, Naitoh Y, Miyata K. The selective 5-hydroxytryptamine (5-HT)₄-receptor agonist RS67506 enhances lower intestinal propulsion in mice. *Jpn J Pharmacol*. 1997;74:209-212.
- Baganz NL, Blakely RD. A dialogue between the immune system and brain, spoken in the language of serotonin. *ACS Chem Neurosci*. 2013;4:48-63.
- Manocha M, Khan WI. Serotonin and GI disorders: an update on clinical and experimental studies. *Clin Transl Gastroenterol*. 2012;3:e13.
- Chen JX, Pan H, Rothman TP, Wade PR, Gershon MD. Guinea pig 5-HT transporter: cloning, expression, distribution, and function in intestinal sensory reception. *Am J Physiol*. 1998;275:G433-G448.
- Chen JJ, Li Z, Pan H, et al. Maintenance of serotonin in the intestinal mucosa and ganglia of mice that lack the high-affinity serotonin transporter: abnormal intestinal motility and the expression of cation transporters. *J Neurosci*. 2001;21:6348-6361.
- Wade PR, Chen J, Jaffe B, Kassem IS, Blakely RD, Gershon MD. Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. *J Neurosci*. 1996;16:2352-2364.
- Borowsky B, Hoffman BJ. Neurotransmitter transporters: molecular biology, function, and regulation. *Int Rev Neurobiol*. 1995;38:139-199.
- Coates MD, Mahoney CR, Linden DR, et al. Molecular defects in mucosal serotonin content and decreased serotonin reuptake transporter in ulcerative colitis and irritable bowel syndrome. *Gastroenterology*. 2004;126:1657-1664.
- Spigset O. Adverse reactions of selective serotonin reuptake inhibitors: reports from a spontaneous reporting system. *Drug Saf*. 1999;20:277-287.
- Gorard DA, Libby GW, Farthing MJ. Influence of antidepressants on whole gut and oro-caecal transit times in health and irritable bowel syndrome. *Aliment Pharmacol Ther*. 1994;8:159-166.
- Gorard DA, Libby GW, Farthing MJ. 5-Hydroxytryptamine and human small intestinal motility: effect of inhibiting 5-hydroxytryptamine reuptake. *Gut*. 1994;35:496-500.

37. Linden DR, White SL, Brooks EM, Mawe GM. Novel promoter and alternate transcription start site of the human serotonin reuptake transporter in intestinal mucosa. *Neurogastroenterol Motil.* 2009;21:e510-e531.
38. Molodecky NA, Soon IS, Rabi DM, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology* 2012;142:46-54 e42; quiz e30.
39. Dahlhamer JM, Zammitti EP, Ward BW, Wheaton AG, Croft JB. Prevalence of inflammatory bowel disease among adults aged ≥ 18 Years – United States, 2015. *Morb Mortal Wkly Rep.* 2016;65:1166-1169.
40. Turner D, Lev-Tzion R. Understanding infliximab in Crohn's disease: the long-term outcomes. *Dig Dis Sci.* 2013;58:604-607.
41. Martinez-Montiel MP, Casis-Herce B, Gomez-Gomez GJ, et al. Pharmacologic therapy for inflammatory bowel disease refractory to steroids. *Clin Exp Gastroenterol.* 2015;8:257-269.
42. Coates MD, Lahoti M, Binion DG, Szigethy EM, Regueiro MD, Bielefeldt K. Abdominal pain in ulcerative colitis. *Inflamm Bowel Dis.* 2013;19:2207-2214.
43. Vivinus-Nebot M, Frin-Mathy G, Bziouche H, et al. Functional bowel symptoms in quiescent inflammatory bowel diseases: role of epithelial barrier disruption and low-grade inflammation. *Gut.* 2014;63:744-752.
44. Click B, Ramos Rivers C, Koutroubakis IE, et al. Demographic and clinical predictors of high healthcare use in patients with inflammatory bowel disease. *Inflamm Bowel Dis.* 2016;22:1442-1449.
45. Oshima S, Fujimura M, Fukimiya M. Changes in number of serotonin-containing cells and serotonin levels in the intestinal mucosa of rats with colitis induced by dextran sodium sulfate. *Histochem Cell Biol.* 1999;112:257-263.
46. Linden DR, Chen JX, Gershon MD, Sharkey KA, Mawe GM. Serotonin availability is increased in mucosa of guinea pigs with TNBS-induced colitis. *Am J Physiol Gastrointest Liver Physiol.* 2003;285:G207-G216.
47. Linden DR, Sharkey KA, Ho W, Mawe GM. Cyclooxygenase-2 contributes to dysmotility and enhanced excitability of myenteric AH neurones in the inflamed guinea pig distal colon. *J Physiol.* 2004;557:191-205.
48. Wheatcroft J, Wakelin D, Smith A, Mahoney CR, Mawe G, Spiller R. Enterochromaffin cell hyperplasia and decreased serotonin transporter in a mouse model of postinfectious bowel dysfunction. *Neurogastroenterol Motil.* 2005;17:863-870.
49. O'Hara JR, Lomax AE, Mawe GM, Sharkey KA. Ileitis alters neuronal and enteroendocrine signalling in guinea pig distal colon. *Gut.* 2007;56:186-194.
50. El-Salhy M, Hatlebakk JG. Changes in enteroendocrine and immune cells following colitis induction by TNBS in rats. *Mol Med Rep.* 2016;14:4967-4974.
51. Magro F, Fraga S, Azevedo I, Soares-da-Silva P. Intestinal 5-hydroxytryptamine and mast cell infiltration in rat experimental colitis. *Dig Dis Sci.* 2006;51:495-501.
52. O'Hara JR, Ho W, Linden DR, Mawe GM, Sharkey KA. Enteroendocrine cells and 5-HT availability are altered in mucosa of guinea pigs with TNBS ileitis. *Am J Physiol Gastrointest Liver Physiol.* 2004;287:G998-G1007.
53. Bailey C, Ruaux C, Stang BV, Valentine BA. Expression of serotonin, chromogranin-A, serotonin receptor-2B, tryptophan hydroxylase-1, and serotonin reuptake transporter in the intestine of dogs with chronic enteropathy. *J Vet Diagn Invest.* 2016;28:271-278.
54. Qian BF, El-Salhy M, Melgar S, Hammarstrom ML, Danielsson A. Neuroendocrine changes in colon of mice with a disrupted IL-2 gene. *Clin Exp Immunol.* 2000;120:424-433.
55. Shajib MS, Wang H, Kim JJ, et al. Interleukin 13 and serotonin: linking the immune and endocrine systems in murine models of intestinal inflammation. *PLoS ONE.* 2013;8:e72774.
56. Magro F, Vieira-Coelho MA, Fraga S, et al. Impaired synthesis or cellular storage of norepinephrine, dopamine, and 5-hydroxytryptamine in human inflammatory bowel disease. *Dig Dis Sci.* 2002;47:216-224.
57. Regmi SC, Park SY, Ku SK, Kim JA. Serotonin regulates innate immune responses of colon epithelial cells through Nox2-derived reactive oxygen species. *Free Radic Biol Med.* 2014;69:377-389.
58. Ghia JE, Li N, Wang H, et al. Serotonin has a key role in pathogenesis of experimental colitis. *Gastroenterology.* 2009;137:1649-1660.
59. Kim JJ, Wang H, Terc JD, Zambrowicz B, Yang QM, Khan WI. Blocking peripheral serotonin synthesis by telotristat etiprate (LX1032/LX1606) reduces severity of both chemical- and infection-induced intestinal inflammation. *Am J Physiol Gastrointest Liver Physiol.* 2015;309:G455-G465.
60. Margolis KG, Stevanovic K, Li Z, et al. Pharmacological reduction of mucosal but not neuronal serotonin opposes inflammation in mouse intestine. *Gut.* 2014;63:928-937.
61. Pascual D, Alsasua A, Goicoechea C, Martin MI. The involvement of 5-HT₃ and 5-HT₄ receptors in two models of gastrointestinal transit in mice. *Neurosci Lett.* 2002;326:163-166.
62. Verity MA, Mellinkoff SM, Frankland M, Greipel M. Serotonin content and argentaffin and Paneth cell changes in ulcerative colitis. *Gastroenterology.* 1962;43:24-31.
63. Capurso L, Friedmann CA. Distribution of 5-OH tryptamine (serotonin) in ulcerative colitis. *Proc R Soc Med.* 1970;63(Suppl.):20-21.
64. El-Salhy M, Danielsson A, Stenling R, Grimelius L. Colonic endocrine cells in inflammatory bowel disease. *J Intern Med.* 1997;242:413-419.
65. Stoyanova II, Gulubova MV. Mast cells and inflammatory mediators in chronic ulcerative colitis. *Acta Histochem.* 2002;104:185-192.
66. Sikander A, Sinha SK, Prasad KK, Rana SV. Association of serotonin transporter promoter polymorphism (5-HTTLPR) with microscopic colitis and ulcerative colitis. *Dig Dis Sci.* 2015;60:887-894.
67. Minderhoud IM, Oldenburg B, Schipper ME, ter Linde JJ, Samsom M. Serotonin synthesis and uptake in symptomatic patients with Crohn's disease in remission. *Clin Gastroenterol Hepatol.* 2007;5:714-720.
68. Shen B, Liu W, Remzi FH, et al. Enterochromaffin cell hyperplasia in irritable pouch syndrome. *Am J Gastroenterol.* 2008;103:2293-2300.
69. Bertrand PP, Barajas-Espinosa A, Neshat S, Bertrand RL, Lomax AE. Analysis of real-time serotonin (5-HT) availability during experimental colitis in mouse. *Am J Physiol Gastrointest Liver Physiol.* 2010;298:G446-G455.
70. Kidd M, Gustafsson BI, Drozdov I, Modlin IM. IL1 β - and LPS-induced serotonin secretion is increased in EC cells derived from Crohn's disease. *Neurogastroenterol Motil.* 2009;21:439-450.
71. Wang Y, Gong H, Lopez R, et al. Correlation between serum serotonin and endoscopy inflammation scores in patients with ileal pouches. *J Crohns Colitis.* 2013;7:e133-e142.
72. Matsumoto K, Lo MW, Hosoya T, et al. Experimental colitis alters expression of 5-HT receptors and transient receptor potential vanilloid 1 leading to visceral hypersensitivity in mice. *Lab Invest.* 2012;92:769-782.
73. Rapalli A, Bertoni S, Arcaro V, et al. Dual role of endogenous serotonin in 2,4,6-trinitrobenzene sulfonic acid-induced colitis. *Front Pharmacol.* 2016;7:68.
74. Nau F Jr, Yu B, Martin D, Nichols CD. Serotonin 5-HT_{2A} receptor activation blocks TNF- α mediated inflammation in vivo. *PLoS ONE.* 2013;8:e75426.
75. Fakhfouri G, Rahimian R, Daneshmand A, et al. Granisetron ameliorates acetic acid-induced colitis in rats. *Hum Exp Toxicol.* 2010;29:321-328.
76. Mousavizadeh K, Rahimian R, Fakhfouri G, Aslani FS, Ghafourifar P. Anti-inflammatory effects of 5-HT receptor antagonist, tropisetron on experimental colitis in rats. *Eur J Clin Invest.* 2009;39:375-383.

77. Belkind-Gerson J, Hotta R, Nagy N, et al. Colitis induces enteric neurogenesis through a 5-HT₄-dependent mechanism. *Inflamm Bowel Dis*. 2015;21:870-878.
78. Spohn SN, Bianco F, Scott RB, et al. Protective actions of epithelial 5-hydroxytryptamine 4 receptors in normal and inflamed colon. *Gastroenterology*. 2016;151:933-944.e933.
79. Matsuyoshi H, Kuniyasu H, Okumura M, et al. A 5-HT₄-receptor activation-induced neural plasticity enhances in vivo reconstructs of enteric nerve circuit insult. *Neurogastroenterol Motil*. 2010;22:806-813.e226.
80. Liu MT, Kuan YH, Wang J, Hen R, Gershon MD. 5-HT₄ receptor-mediated neuroprotection and neurogenesis in the enteric nervous system of adult mice. *J Neurosci*. 2009;29:9683-9699.
81. Kim JJ, Bridle BW, Ghia JE, et al. Targeted inhibition of serotonin type 7 (5-HT₇) receptor function modulates immune responses and reduces the severity of intestinal inflammation. *J Immunol*. 2013;190:4795-4804.
82. Guseva D, Holst K, Kaune B, et al. Serotonin 5-HT₇ receptor is critically involved in acute and chronic inflammation of the gastrointestinal tract. *Inflamm Bowel Dis*. 2014;20:1516-1529.
83. Yu FY, Huang SG, Zhang HY, et al. Comparison of 5-hydroxytryptophan signaling pathway characteristics in diarrhea-predominant irritable bowel syndrome and ulcerative colitis. *World J Gastroenterol*. 2016;22:3451-3459.
84. Linden DR, Foley KF, McQuoid C, Simpson J, Sharkey KA, Mawe GM. Serotonin transporter function and expression are reduced in mice with TNBS-induced colitis. *Neurogastroenterol Motil*. 2005;17:565-574.
85. Tada Y, Ishihara S, Kawashima K, et al. Downregulation of serotonin reuptake transporter gene expression in healing colonic mucosa in presence of remaining low-grade inflammation in ulcerative colitis. *J Gastroenterol Hepatol*. 2016;31:1443-1452.
86. Bertrand PP, Bertrand RL, Camello PJ, Pozo MJ. Simultaneous measurement of serotonin and melatonin from the intestine of old mice: the effects of daily melatonin supplementation. *J Pineal Res*. 2010;49:23-34.
87. Koh SJ, Kim JM, Kim IK, et al. Fluoxetine inhibits NF-kappaB signaling in intestinal epithelial cells and ameliorates experimental colitis and colitis-associated colon cancer in mice. *Am J Physiol Gastrointest Liver Physiol*. 2011;301:G9-G19.
88. Minaiyan M, Hajhashemi V, Rabbani M, Fattahian E, Mahzouni P. Evaluation of anti-colitic effect of flvoxamine against acetic acid-induced colitis in normal and reserpinized depressed rats. *Eur J Pharmacol*. 2015;746:293-300.
89. Koh SJ, Kim JW, Kim BG, Lee KL, Im JP, Kim JS. Fluoxetine inhibits hyperresponsive lamina propria mononuclear cells and bone marrow-derived dendritic cells, and ameliorates chronic colitis in IL-10-deficient mice. *Dig Dis Sci*. 2015;60:101-108.
90. Haub S, Ritze Y, Bergheim I, Pabst O, Gershon MD, Bischoff SC. Enhancement of intestinal inflammation in mice lacking interleukin 10 by deletion of the serotonin reuptake transporter. *Neurogastroenterol Motil*. 2010;22:826-834.e229.
91. Bischoff SC, Mailer R, Pabst O, et al. Role of serotonin in intestinal inflammation: knockout of serotonin reuptake transporter exacerbates 2,4,6-trinitrobenzene sulfonic acid colitis in mice. *Am J Physiol Gastrointest Liver Physiol*. 2009;296:G685-G695.
92. Costedio MM, Coates MD, Danielson AB, et al. Serotonin signaling in diverticular disease. *J Gastrointest Surg*. 2008;12:1439-1445.
93. Foley KF, Pantano C, Ciolino A, Mawe GM. IFN-gamma and TNF-alpha decrease serotonin transporter function and expression in Caco2 cells. *Am J Physiol Gastrointest Liver Physiol*. 2007;292:G779-G784.
94. Latorre E, Mendoza C, Matheus N, et al. IL-10 modulates serotonin transporter activity and molecular expression in intestinal epithelial cells. *Cytokine*. 2013;61:778-784.
95. Macer BJ, Prady SL, Mikocka-Walus A. Antidepressants in inflammatory bowel disease: a systematic review. *Inflamm Bowel Dis*. 2017;23:534-550.
96. Mikocka-Walus AA, Turnbull DA, Moulding NT, Wilson IG, Andrews JM, Holtmann GJ. Antidepressants and inflammatory bowel disease: a systematic review. *Clin Pract Epidemiol Ment Health*. 2006;2:24.
97. Bonderup OK, Fenger-Gron M, Wigh T, Pedersen L, Nielsen GL. Drug exposure and risk of microscopic colitis: a nationwide Danish case-control study with 5751 cases. *Inflamm Bowel Dis*. 2014;20:1702-1707.
98. Fernandez-Banares F, Esteve M, Espinos JC, et al. Drug consumption and the risk of microscopic colitis. *Am J Gastroenterol*. 2007;102:324-330.
99. Taylor JK. Retroperitoneal fibrosis, regional enteritis, and carcinoid tumors. *JAMA*. 1971;217:1864.
100. Hsu EY, Feldman JM, Lichtenstein GR. Ileal carcinoid tumors stimulating Crohn's disease: incidence among 176 consecutive cases of ileal carcinoid. *Am J Gastroenterol*. 1997;92:2062-2065.
101. Vera-Portocarrero LP, Ossipov MH, King T, Porreca F. Reversal of inflammatory and noninflammatory visceral pain by central or peripheral actions of sumatriptan. *Gastroenterology*. 2008;135:1369-1378.
102. Rahimi R, Nikfar S, Abdollahi M. Efficacy and tolerability of *Hypericum perforatum* in major depressive disorder in comparison with selective serotonin reuptake inhibitors: a meta-analysis. *Prog Neuropsychopharmacol Biol Psychiatry*. 2009;33:118-127.
103. Ford AC, Quigley EM, Lacy BE, et al. Effect of antidepressants and psychological therapies, including hypnotherapy, in irritable bowel syndrome: systematic review and meta-analysis. *Am J Gastroenterol*. 2014;109:1350-1365; quiz 1366.
104. Chang L, Lembo A, Sultan S. American Gastroenterological Association Institute Technical Review on the pharmacological management of irritable bowel syndrome. *Gastroenterology*. 2014;147:1149-1172 e1142.
105. Coates MD, Johnson AC, Greenwood-Van Meerveld B, Mawe GM. Effects of serotonin transporter inhibition on gastrointestinal motility and colonic sensitivity in the mouse. *Neurogastroenterol Motil*. 2006;18:464-471.
106. Muller-Lissner SA, Fumagalli I, Bardhan KD, et al. Tegaserod, a 5-HT₄ receptor partial agonist, relieves symptoms in irritable bowel syndrome patients with abdominal pain, bloating and constipation. *Aliment Pharmacol Ther*. 2001;15:1655-1666.
107. Hoffman JM, Tyler K, MacEachern SJ, et al. Activation of colonic mucosal 5-HT₄ receptors accelerates propulsive motility and inhibits visceral hypersensitivity. *Gastroenterology*. 2012;142:844-854 e844.

How to cite this article: Coates MD, Tekin I, Vrana KE, Mawe GM. Review article: the many potential roles of intestinal serotonin (5-hydroxytryptamine, 5-HT) signalling in inflammatory bowel disease. *Aliment Pharmacol Ther*. 2017;46:569-580. <https://doi.org/10.1111/apt.14226>